DIMERS OF ISOEUGENOL BY DYE-SENSITIZED PHOTOOXIDATION

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The oxidation of isoeugenol to a variety of compounds is well documented in the literature (1-5). We have extended this research by examining the effect of dye-sensitized photooxidation on isoeugenol (I), a monomer that has been implicated in the biosynthesis of lignin (6).

Isolation of "lignol-like" dimers from the photomixture gives reason to consider the possibility that photooxidation may contribute to lignin synthesis.

In a typical reaction, 1 g (6.10 mmole) of cis-trans isoeugenol and 25 mg of protoporphyrin dihydrochloride were dissolved in 100 ml of methanol. The solution, constantly saturated with a stream of oxygen, was photooxidized by two 300-W incandescent lamps for 24 hr. The ether solution of the photomixture, after extraction with water, yielded 1040 mg of crude photo-product. Blank runs under nitrogen or under oxygen without any dye gave essentially no reaction. Photooxidation of isoeugenol acetate under the same conditions also produced no products. TLC and column chromatography (silica gel, cyclohexane-ether mixtures) showed the presence of, and partially separated, three major photoproducts--A, B, and C. Separation of A and C was completed by acetylation of the photomixture before column chromatography. In

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one run, 540 mg of crude acetylated photomixture gave 95 mg of acetylated A, 110 mg of acetylated B, and 115 mg of acetylated C.

Compounds B and C were catalytically hydrogenated (Pd, C, 1 atm) to eliminate the occurrence of cis,trans isomers. NMR data for compounds B and C, therefore, are given only for the acetylated and hydrogenated compounds.

Major products--A, B, and C--were identified by a combination of chemical and instrumental techniques. Compound A was identified as dehydrodiisoeugenol by comparison of its melting point and that of its p-nitrobenzoyl derivative with reported values.

M.p. 153-5 (cf. 6), p-nitrobenzoyl ester m.n. 155-7 (cf. 6)
λ max (H2O) 274 μm
Calc.: C, 75.62; H, 6.75
Found: C, 75.72; H, 6.83
ms. m/z M+ 326 (C20H22O4)

Compound A, dehydrodiisoeugenol

Elemental analysis and high resolution mass spectroscopy both confirmed the molecular formula C20H22O4. NMR data for A 1.36(3H,d), 1.85(3H,d), 3.44(1H,m), 3.84(3H,s), 3.56(3H,s), 5.08(1H,d), 5.58(1H,s), 6.10(2H,m), 6.82(2H,m) and hydrogenated, acetylated A corresponded to the proposed structure.

Compounds B and C were separated and identified as their acetates. Elemental analysis and mass spectroscopy gave a molecular formula of C23H28O6 for both acetylated B and C. The mass spectra for both acetylated B and C were identical. Catalytic hydrogenation of acetylated B and C gave two compounds, which also fragmented in identical patterns. These fragmentation patterns (Table I), together with elemental analysis and NMR data (Table II), strongly suggested the following structure for both compounds:
B and C (acetylated) 1 methoxy-1-[4-acetoxy-3-methoxyphenyl]-2-[4-propenyl-2-methoxyphenoxy] propane

TABLE I

Major Mass Spectral Lines for Compounds B and C

<table>
<thead>
<tr>
<th>m/e</th>
<th>402</th>
<th>237</th>
<th>236</th>
<th>209</th>
<th>195</th>
<th>194</th>
<th>193</th>
<th>167</th>
<th>166</th>
<th>137</th>
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</thead>
<tbody>
<tr>
<td>Intensity</td>
<td>24</td>
<td>24</td>
<td>18</td>
<td>22</td>
<td>56</td>
<td>40</td>
<td>46</td>
<td>100</td>
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<table>
<thead>
<tr>
<th>m/e</th>
<th>400</th>
<th>237</th>
<th>236</th>
<th>209</th>
<th>195</th>
<th>194</th>
<th>191</th>
<th>167</th>
<th>164</th>
<th>137</th>
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</thead>
<tbody>
<tr>
<td>Intensity</td>
<td>28</td>
<td>25</td>
<td>28</td>
<td>9</td>
<td>81</td>
<td>51</td>
<td>21</td>
<td>100</td>
<td>30</td>
<td>18</td>
</tr>
</tbody>
</table>

a Acetylated, hydrogenated.

b Acetylated.

Differences in NMR and in chromatography between B and C may be explained by the diastereomeric forms possible. The NMR spectra of both B and C fit the proposed general structure well, but it was not possible to fit either one to a specific diastereomeric form. It was also difficult to explain the large shift in the saturated methyl absorption (b)
from 1.30 in compound B to 1.09 in compound C. Conformational differences may be responsible, but they are not readily apparent.

**TABLE II**

<table>
<thead>
<tr>
<th>B</th>
<th>C</th>
<th>B</th>
<th>C</th>
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</thead>
<tbody>
<tr>
<td>(a)</td>
<td>0.96(3H,t)</td>
<td>0.92(3H,t)</td>
<td>(f)</td>
</tr>
<tr>
<td>(b)</td>
<td>1.30(3H,d)</td>
<td>1.09(3H,d)</td>
<td>(g)</td>
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<tr>
<td>(c)</td>
<td>1.58(2H,m)</td>
<td>1.61(2H,m)</td>
<td>(h)</td>
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<tr>
<td>(d)</td>
<td>2.27(3H,s)</td>
<td>2.28(3H,s)</td>
<td>(i)</td>
</tr>
<tr>
<td>(e)</td>
<td>2.49(2H,t)</td>
<td>2.51(2H,t)</td>
<td>(j)</td>
</tr>
</tbody>
</table>

* Acetylated and hydrogenated.

Formation of dehydrodiisoeugenol and the isomeric S-aryl ethers B and C by dye-sensitized photooxidation suggest that lignin formation could be initiated by a light-dye-O₂ system in the plant. A possible mechanism involves the abstraction of the phenolic hydrogen by an excited dye triplet or singlet oxygen to produce the products via a radical addition involving a quinone methide radical intermediate (7). Recent work by Knowles (8) has implicated the triplet state of flavins in the photooxidation of nucleotides. A number of other workers (9,10) have shown the hydrogen abstracting abilities of singlet oxygen. It is possible that plant pigments and oxygen may function similarly in lignin synthesis.

REFERENCES