Stocking strategies for production of *Litopenaeus vannamei* (Boone) in amended freshwater in inland ponds

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Abstract

The performance of the Pacific white shrimp *Litopenaeus vannamei* (Boone) under various stocking strategies was evaluated in earthen ponds filled with freshwater amended with major ions. Six 0.1-ha earthen ponds located in Pine Bluff, AR, USA, were filled with freshwater in 2003 and 2004, and potassium magnesium sulphate added to provide 50 mg K\(^+\) L\(^-1\) and stock salt added to provide 0.5 g L\(^-1\) salinity. In 2003, three ponds either were stocked with PL\(_{15}\) shrimp (39 PL m\(^2\)) for 125 days of grow out or with PL\(_{25}\) shrimp for 55 days (23 PL m\(^2\)) followed by a 65-day (28 PL m\(^2\)) grow-out period. In 2004, ponds were stocked with 7, 13 or 30 PL\(_{15}\) m\(^2\) for 134 days of grow out. Salinity averaged 0.7 g L\(^-1\) during both years, and concentration of SO\(_4\)\(^2-\), K\(^+\), Ca\(^2+\) and Mg\(^2+\) was higher, and Na\(^+\) and Cl\(^-\) was lower in amended pond water than in seawater at 0.7 g L\(^-1\) salinity. Potassium concentration in amended water was 52–81% of the target concentration. Shrimp yields ranged from 3449 kg ha\(^-1\) in 2003 to 4966 kg ha\(^-1\) in 2004 in ponds stocked with 30–39 PL\(_{15}\) m\(^2\) for a 125–134-day culture period. At harvest, mean individual weight ranged from 17.1 to 19.3 g shrimp\(^-1\). In ponds stocked with PL\(_{25}\) shrimp, yields averaged 988 and 2462 kg ha\(^-1\) for the 1st and 2nd grow-out periods respectively. Gross shrimp yield increased in 2004 increased linearly from 1379–4966 kg ha\(^-1\) with increased stocking rate. These experiments demonstrated that *L. vannamei* can be grown successfully in freshwater supplemented with major ions to a final salinity of 0.7 g L\(^-1\).

Keywords: *Litopenaeus vannamei* (Boone), freshwater, mineral amendment, aquaculture, inland ponds

Introduction

The Pacific white shrimp *Litopenaeus vannamei* (Boone) is being cultured far from coastal areas in ponds filled with low-salinity (generally 2–5 g L\(^-1\)) ground water or brine transported from the coast and diluted on site with freshwater. Six ions in seawater comprise 99.8%, by weight, of salinity: Cl\(^-\) (55.3%), Na\(^+\) (30.8%), SO\(_4\)\(^2-\) (7.7%), Mg\(^2+\) (3.7%), Ca\(^2+\) (1.2%), and K\(^+\) (1.1%). The ionic composition of low-salinity ground water varies geographically and often differs from that of dilute seawater at the same salinity. Boyd and Thunjai (2003) and Saoud, Davis and Rouse (2003) collected water samples for ion analysis from inland shrimp ponds and wells in Alabama, Arizona, Florida, Mississippi and Texas in the United States and in Australia, China, Ecuador and Thailand. Although ion concentrations in many samples were similar to those of dilute seawater at the same salinity, there were some notable deviations. Deficiencies in chloride, magnesium, potassium and sulphate were observed in some samples, while excesses of calcium and sulphate were observed in other samples. Relative to dilute seawater at the same salinity, ion deficiencies were most common in ponds filled with low-salinity ground water.

The availability of brackish ground water in non-coastal regions provides an opportunity to culture...
marine species able to adapt to reduced salinity. Ratios of major ions in low-salinity ground water may serve as indicators of the suitability of brackish ground water for aquaculture. Among the ionic ratios that have been evaluated in relation to survival and growth of marine species are Na\(^+\):K\(^+\) and K\(^+\):Cl\(^-\) for red drum (Sciaenops ocellatus [Linneaus]) (Forsberg, Dorsett & Neill 1996), K\(^+\):Cl\(^-\) for Australian snapper (Pagrus auratus [Forster]) (Fielder, Bardsley & Allan 2001), Na\(^+\):Ca\(^{2+}\) (Atwood, Young, Tomasso & Browdy 2003) and Na\(^+\):K\(^+\) (Zhu, Dong, Wang & Huang 2004) for L. vannamei. These ratios also can be used to develop remediation strategies to correct ionic imbalances.

Ionic imbalance in inland saline ground water can affect shrimp survival. Saoud et al. (2003) reported variable L. vannamei survival 48 h after initiating acclimation to 0.7–16.3 g L\(^{-1}\) salinity well waters from various sources. Farmers in the southern United States have reported poor growth and survival for L. vannamei stocked in inland ponds filled with low-salinity water deficient in one or more ions. Addition of specific fertilizers to supply or augment concentrations of specific deficient ions appeared to solve the problem. Boyd and Thunjai (2003) and McNevin, Boyd, Silapajarn and Silapajarn (2004) recommended a number of minerals that could be used to supplement major cations in low-salinity water.

Because low-salinity ground water can be amended to supply one or more deficient ions, it should be possible to amend water from freshwater (< 0.5 g L\(^{-1}\) salinity) aquifers to support successful shrimp culture. Results of a preliminary trial conducted in 2002 showed that L. vannamei survived and grew in freshwater supplemented with major ions (B.W. Green, unpublished data). In that trial, a target of 50 mg K\(^+\) L\(^{-1}\) was selected to ensure adequate soluble K\(^+\) in pond water because K\(^+\) can be adsorbed strongly by clay soils (Bohn, McNeal & O’Connor 1985). Furthermore, similar levels of K\(^+\) supplementation were practiced in Alabama in response to shrimp mortality putatively caused by K\(^+\) deficiency (McNevin et al. 2004). Potassium magnesium sulphate fertilizer, selected because it provides three major ions, was applied to provide 50 mg K\(^+\) L\(^{-1}\). Inland culture in the United States, shrimp 17.5–21.0 g shrimp\(^{-1}\) and larger are considered marketable (Samocha, Hamper, Emberson, Davis, McIntosh, Lawrence & VanWyk 2002). Thus, the objective of the present research was to evaluate the performance of L. vannamei under different stocking strategies for producing marketable shrimp in earthen ponds filled with freshwater amended with major ions.

**Materials and methods**

Trials were conducted in 2003 and 2004. In both trials, six 0.1-ha earthen ponds located on the Aquaculture Research Station, University of Arkansas at Pine Bluff, Pine Bluff, AR, were filled with freshwater (total alkalinity 126 mg L\(^{-1}\) as CaCO\(_3\), total hardness 162 mg L\(^{-1}\) as CaCO\(_3\)) from a 2.0-ha reservoir. Water to fill the reservoir was pumped from a 61-m-deep well. Pond water levels were maintained approximately 15 cm below the top of the standpipe to capture rainfall. Reservoir water was added as needed to maintain water level in ponds 15–30 cm below the top of the standpipe. No water was exchanged in ponds. Each pond was equipped with a 0.37-kW electric paddlewheel aerator (Southern Machine Welding, Quinton, AL, USA) that was operated nightly during each trial.

Before flooding agricultural limestone (250 mesh) was spread on pond bottoms (Table 1). Livestock salt (ca. 96% NaCl) and potassium magnesium sulphate fertilizer (K-MAG Standard guaranteed analysis: 22.0% K\(_2\)O, 10.8% Mg, 22.0% S, The Mosaic Company, Plymouth, MN, USA) (Table 1) was added to ponds generally to increase concentrations of the six major ions found in seawater and specifically to increase K\(^+\) concentration. Target concentrations selected were 50 mg K\(^+\) L\(^{-1}\) from potassium magnesium sulphate and 0.5 g L\(^{-1}\) of salinity from the livestock salt. Input quantities were reduced in 2004 based on 2003 water analysis results and improved pond volume estimates. The potassium magnesium sulphate was dissolved in pond water in buckets before being applied to the pond. During the first 21 days of both years, two of the six ponds lost water to seepage and received an additional 1130–1361 kg ha\(^{-1}\) of livestock salt to replace salinity lost to dilution.

**Table 1** Quantities of agricultural limestone (250 mesh), livestock salt (ca. 96% NaCl), and potassium magnesium sulphate (0-0-22 11% Mg, 22% S) added to inland freshwater ponds to increase mean salinity to 0.7 g L\(^{-1}\)

<table>
<thead>
<tr>
<th>Trial year</th>
<th>Agricultural limestone (kg ha(^{-1}))</th>
<th>Livestock salt (kg ha(^{-1}))</th>
<th>Potassium magnesium sulphate fertilizer (kg ha(^{-1}))</th>
</tr>
</thead>
<tbody>
<tr>
<td>2003</td>
<td>1100</td>
<td>5670</td>
<td>3175</td>
</tr>
<tr>
<td>2004</td>
<td>567</td>
<td>4536</td>
<td>2722</td>
</tr>
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</table>
Water samples were collected in 2003 from each of three randomly selected ponds before and from all ponds 1 day after salinity adjustment and from three ponds after 125 days of continuous culture. The Water Quality Laboratory, Arkansas Water Resources Center, University of Arkansas, Fayetteville, AR, USA, analysed all water samples for fluoride, chloride and sulphate using ion chromatography (IC), and calcium, magnesium, potassium and sodium using inductively coupled plasma spectroscopy (ICP). In addition, two of the day-125 samples were analysed for aluminum, cobalt, copper, iron, manganese, molybdenum and zinc by ICP. In 2004, two samples of pipe water discharging into ponds during pond flooding were collected for ion analysis. Water samples were collected from all ponds on days 5, 105, and 134. All 2004 water samples were analysed on-site using IC for calcium, chloride, magnesium, potassium, sodium and sulphate.

In 2003, each pond was fertilized with cottonseed meal and chemical fertilizer (19-19-19) on days 1 and 6 after filling to stimulate natural productivity. A total of 225 kg ha\(^{-1}\) cottonseed meal and 30 kg ha\(^{-1}\) 19-19-19 were added. In 2004, each pond was fertilized to stimulate natural productivity 1 day after filling with cottonseed meal (227 kg ha\(^{-1}\)), wheat bran (227 kg ha\(^{-1}\)) and minnow meal (73 kg ha\(^{-1}\)). Each pond was fertilized (38.6 kg ha\(^{-1}\), 19-19-19) on days 1, 7 and 14. Chemical fertilizer was dissolved in pond water in a bucket before being applied to the pond.

Postlarval (PL\(_{15}\) and PL\(_{25}\)) *L. vannamei* (BP101 line), purchased both years from the GMSB Shrimp Hatchery in Florida, USA, were held in tanks for 56 h while acclimated to pond salinity. Shrimp were fed during acclimation with a 50% protein feed (850–1200 \(\mu\)m particle size; PL Raceway Plus, Zeigler Brothers). At stocking, average individual PL weight was determined by dividing the bulk weight of PLs in a tared beaker of water by the number of PLs. Numbers at stocking were determined gravimetrically.

A completely randomized design was used for both trials. Producing one or two crops in a single growing season was tested in 2003. The planned stocking rate (33 PL m\(^{-2}\)) was similar to that used at inland farms; however, PL availability following acclimation resulted in deviation from the plan. Three ponds were stocked on 30 May 2003 with 39 PL\(_{15}\) shrimp m\(^{-2}\) and grown for the entire growing season (125 days). The other three ponds were stocked the same day with 23 PL\(_{25}\) shrimp m\(^{-2}\), grown for 55 days, and harvested. Ponds were re-flooded, salinity adjusted to 0.7–0.8 g l\(^{-1}\), and 28 PL\(_{25}\) shrimp m\(^{-2}\) were stocked on 1 August 2003 and grown for 65 days. The effect on growth and yield of stocking 7, 13 or 30 PL\(_{15}\) m\(^{-2}\) was tested in 2004. On 8 May 2004 ponds in the 7 and 13 PL\(_{25}\) m\(^{-2}\) treatments were stocked completely. Ponds in the 30 PL\(_{15}\) m\(^{-2}\) treatment were stocked partially (60%) because of insufficient PLs; stocking was completed on 15 May 2004 upon receipt of a second PL shipment. Ponds were harvested on 20–21 September 2004.

Shrimp in all ponds were fed a commercially formulated, sinking, extruded shrimp diet (35% crude protein, Shrimp Grow-Out SI-35, Zeigler Brothers), 7 days week\(^{-1}\). A constant quantity of feed (22.7 and 13.6 kg ha\(^{-1}\) in 2003 and 2004 respectively) was added to ponds daily for the first 27–33 days. Beginning days 28–34, feeding rate was adjusted weekly, decreasing from 8.2% to 1.8% of the biomass as shrimp grew (Clifford III 1992). The shrimp population in each pond was sampled weekly beginning days 28–34 to monitor growth. Shrimp (20–25 shrimp pond\(^{-1}\)) were captured using a cast net, weighed individually to the nearest 0.1 g, and returned to the pond. Specific growth rate (SGR) of shrimp was calculated: 

\[
SGR = 100 \times \left( \frac{\ln W_t - \ln W_0}{T_t - T_0} \right)
\]

where \(\ln W\) = natural logarithm of mean individual weight (g shrimp\(^{-1}\)) at time \(t\) or \(t+1\) or \(t\), and \(T = \text{time (d)} \times \text{t+1 or } t\). Shrimp were harvested by draining the pond. The total weight of shrimp harvested per pond was measured. A sample of 100 shrimp pond\(^{-1}\) was weighed individually to the nearest 0.1 g. Total number of shrimp harvested per pond was calculated by dividing the total biomass harvested by the mean individual weight. Gross feed conversion ratio was calculated for each replicate by dividing the total weight of feed offered by the total weight of shrimp harvested.

Dissolved oxygen concentration and water temperature were measured daily in ponds between 07:00–08:00 hours, and periodically between 14:00 and 15:00 hours using a YSI oxygen meter (Model 550A, Yellow Springs Instrument, Yellow Springs, OH, USA). Salinity was measured in ponds every 12 days on an average in 2003 and daily in 2004 using a YSI salinity meter (Model 30, Yellow Springs Instrument).

Production data for 2003 were analysed using PROC GLM and regression analysis; percent data were arcsin transformed before analysis (Sokal & Rohlf 1995). Water analyses data for 2003 and 2004 were analysed using a statistical model for repeated measures in the mixed models procedure (PROC MIXED). Data analyses were performed using SAS.
Net feed conversion ratio was calculated by dividing the total quantity of feed fed by the net yield. Differences were declared significant at α level 0.05.

Results

Salinity of pond water before ionic amendment was 0.15–0.20 g L⁻¹. Mineral addition increased mean concentration of major cations and anions (Table 2). Mean pond salinity in both years increased to 0.7–0.9 g L⁻¹ immediately following ionic amendment and averaged 0.7 g L⁻¹ for each grow out cycle. Compared with seawater at 0.7 g L⁻¹ salinity, mean ion concentrations in amended water were higher for calcium, magnesium and potassium, and lower for chloride and sodium.

The effect of ion amendment of major ions persisted throughout the culture period during both years (Table 2). Maximum mean ion concentrations were observed in water samples collected on day 1 (2003) and day 5 (2004). During both years, mean concentration of calcium, magnesium and potassium at the end of the culture period exceeded their corresponding concentration in seawater at 0.7 g L⁻¹ salinity. Mean sodium and chloride concentration at the end of the culture period during both years was less than their corresponding concentration in seawater at 0.7 g L⁻¹ salinity.

Potassium concentration in amended water failed to attain the target of 50 mg K⁺ L⁻¹ in either year (Table 2).

The Na⁺:K⁺ and Ca²⁺:Mg²⁺ ratios of pond water decreased following addition of salts and then increased slightly during both years (Table 2). The Na⁺:Ca²⁺ ratio increased slightly following fertilization, then decreased. The K⁺:Cl⁻ ratio increased following fertilization and throughout the 125-day culture period in 2003, whereas the ratio decreased throughout the culture period in 2004. In comparison, the Na⁺:Ca²⁺, K⁺:Cl⁻ and Ca²⁺:Mg²⁺ ratios in 0.7-g L⁻¹ seawater were higher, higher, lower and lower, respectively, than in the amended pond water.

Table 2   Mean concentration of ions (mg L⁻¹) in 2003 and 2004 in source water and in pond water at various times after mineral amendment. Salinity in ponds during both years averaged 0.7 g L⁻¹ following amendment*

<table>
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</thead>
<tbody>
<tr>
<td>Cl⁻</td>
<td>33.8b</td>
<td>239.0a</td>
<td>183.2b</td>
<td>26.8d</td>
<td>349.6a</td>
<td>253.8b</td>
<td>207.5c</td>
<td>393.7</td>
</tr>
<tr>
<td>SO₄²⁻</td>
<td>36.5b</td>
<td>136.1a</td>
<td>71.7ab</td>
<td>12.1b</td>
<td>54.2a</td>
<td>21.3b</td>
<td>23.0b</td>
<td>55.2</td>
</tr>
<tr>
<td>Na⁺</td>
<td>56.6b</td>
<td>161.4a</td>
<td>197.1a</td>
<td>45.9f</td>
<td>242.5a</td>
<td>188.9b</td>
<td>170.4c</td>
<td>219.1</td>
</tr>
<tr>
<td>K⁺</td>
<td>3.8b</td>
<td>28.1a</td>
<td>28.6b</td>
<td>2.6b</td>
<td>32.1a</td>
<td>18.9b</td>
<td>15.6b</td>
<td>8.1</td>
</tr>
<tr>
<td>Ca²⁺</td>
<td>27.4a</td>
<td>38.9a</td>
<td>54.9a</td>
<td>21.3b</td>
<td>43.5a</td>
<td>37.0b</td>
<td>37.2b</td>
<td>8.4</td>
</tr>
<tr>
<td>Mg²⁺</td>
<td>17.7b</td>
<td>33.2ab</td>
<td>45.6b</td>
<td>19.1b</td>
<td>40.0a</td>
<td>29.1b</td>
<td>26.8b</td>
<td>26.2</td>
</tr>
<tr>
<td>Na⁺:K⁺</td>
<td>25.1</td>
<td>9.8</td>
<td>11.7</td>
<td>26.9</td>
<td>12.8</td>
<td>17.0</td>
<td>18.6</td>
<td>45.9</td>
</tr>
<tr>
<td>Na⁺:Ca²⁺</td>
<td>3.6</td>
<td>7.2</td>
<td>6.3</td>
<td>3.7</td>
<td>9.7</td>
<td>8.9</td>
<td>8.0</td>
<td>45.6</td>
</tr>
<tr>
<td>K⁺:Cl⁻</td>
<td>0.102</td>
<td>0.107</td>
<td>0.142</td>
<td>0.089</td>
<td>0.083</td>
<td>0.067</td>
<td>0.068</td>
<td>0.019</td>
</tr>
<tr>
<td>Ca²⁺:Mg²⁺</td>
<td>0.93</td>
<td>0.71</td>
<td>0.73</td>
<td>0.68</td>
<td>0.66</td>
<td>0.77</td>
<td>0.84</td>
<td>0.19</td>
</tr>
</tbody>
</table>

Concentration of ions in 0.7-g L⁻¹ seawater is shown for comparison. Molar ratios of Na⁺:K⁺, Na⁺:Ca²⁺, K⁺:Cl⁻ and Ca²⁺:Mg²⁺ are shown for each water type.*

*In each row means within year with different superscripts are significantly different (P<0.05).

Table 3   Mean early morning water temperature and dissolved oxygen concentration in treatment ponds during the 2003 and 2004 experiments*

<table>
<thead>
<tr>
<th></th>
<th>2003</th>
<th>2004</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>PL₂₅</td>
<td>PL₂₅⁻₁</td>
</tr>
<tr>
<td>Water temperature (°C)</td>
<td>27.2a</td>
<td>27.9a</td>
</tr>
<tr>
<td>Dissolved oxygen (mg L⁻¹)</td>
<td>4.3b</td>
<td>4.6b</td>
</tr>
</tbody>
</table>

*In each row means within year with different superscripts are significantly different (P<0.05).
Mean concentrations of minor elements measured in the day 125 samples in 2003 were: aluminum $^0.051$ mg L$^{-1}$, cobalt $^0.000$ mg L$^{-1}$, copper $^0.128$ mg L$^{-1}$, iron $^0.041$ mg L$^{-1}$, manganese $^0.363$ mg L$^{-1}$, molybdenum $^0.008$ mg L$^{-1}$, and zinc $^0.052$ mg L$^{-1}$.

Water temperature increased during May and June, remained relatively constant during July and August, and declined during September. Mean early morning water temperatures and dissolved oxygen concentrations are shown by treatment for each year in Table 3.

Gross yield of heads-on shrimp in 2003 was significantly greater in ponds stocked with PL15 shrimp than in ponds stocked with PL25 shrimp (Table 4). However, the sum of gross yield for the first- and second-production cycles where PL25 shrimp were stocked, i.e., the entire growing season, did not differ significantly from the gross yield for PL15 shrimp. Shrimp growth in the PL25 first-cycle treatment did not appear to differ from that in the PL15 treatment (Fig. 1).

Two separate aerator failures during the 2004 experiment resulted in shrimp mass-mortality by hypoxia. The first event occurred on 10 August 2004 (day 94) and the second event on 15 September 2004 (day 130); production data from these ponds were excluded from analyses. Mean production data by treatment are presented in Table 4. Gross yield of heads-on shrimp increased linearly with increased stocking rate ($R^2 = 0.989$; Table 4). Shrimp growth in the 13 and 30 PL m$^{-2}$ treatments appeared to be similar and lower than in the 7 PL m$^{-2}$ treatment (Fig. 1).

**Discussion**

*Litopenaeus vannamei* survived and grew successfully during two consecutive seasons in inland ponds filled with a hard freshwater amended with sodium, chloride, potassium, magnesium and sulphate to a final mean salinity of 0.7 g L$^{-1}$ in the current study.
These results contrast with Laramore, Laramore and Scarpa (2001) who found in a 40-day experiment that PL25 and PL40 L. vannamei did not survive at salinity < 2 g L\(^{-1}\) and had reduced survival and growth at salinity < 4 g L\(^{-1}\). Shrimp survival was significantly lower in water whose source of salinity (2–5 g L\(^{-1}\)) was mixed salts (chlorides of calcium, magnesium, potassium and sodium) compared with sea salt (artificial seawater salt) (Atwood et al. 2003; Sowers, Gatlin, Young, Isely, Brown & Tomasso 2005). However, inclusion of 1 g L\(^{-1}\) salinity from sea salt along with salinity derived from mixed salts improved L. vannamei growth and survival (Atwood et al. 2003; Sowers, Tomasso Jr, Brown & Atwood 2006). Thus, these authors concluded that sea salt is required for growth and survival of L. vannamei.

Low-salinity ground water is used to culture L. vannamei at inland sites in a number of states in the southern United States. Brackish ground water also has been evaluated for inland culture of marine fish such as red drum (Forsberg et al. 1996) or Australian snapper (Fielder et al. 2001). The ionic proportions found in brackish ground water often differ from those in seawater at the same salinity (Forsberg et al. 1996; Fielder et al. 2001; Boyd & Thunjai 2003; Partridge & Creeper 2004). Potassium, magnesium, chloride, and sulphate concentrations in low-salinity ground water can be lower than those in dilute seawater of the salinity (Boyd & Thunjai 2003; Zhu et al. 2004). Inland shrimp farmers in the United States have reported shrimp mortality in low-salinity water whose ionic proportions differed from those in seawater (McNevin et al. 2004). Supplementation of ion-deficient saline groundwater with Mg\(^{2+}\) and/or K\(^{+}\) significantly improved L. vannamei PL17 survival compared with untreated water (Davis, Boyd, Rouse & Saoud 2005). Similarly, red drum survival was increased significantly by increasing concentration of deficient ions by mixed salt addition (Forsberg et al. 1996).

Application of potassium magnesium sulphate increased pond K\(^{+}\), Mg\(^{2+}\) and SO\(_4\)\(^{2-}\) concentrations, but the post-application K\(^{+}\) concentration was only 56–64% of the target concentration. In a small research pond trial, McNevin et al. (2004) noted that the increase in ionic concentrations only was about 66% of the expected increases if the applied potassium magnesium sulphate dissolved completely. Pond soils in inland shrimp ponds adsorbed 55% of added potassium (Boyd, Boyd & Rouse 2007a). Adsorbed potassium was comprised of exchangeable K\(^{+}\) and fixed K\(^{+}\) (Boyd et al. 2007a; Boyd, Boyd & Rouse 2007b). Soil adsorption of K\(^{+}\) likely was responsible for the difference in observed and expected pond water K\(^{+}\) concentration in the present experiment.

In evaluating the initial quality and impact of ion amendment to correct deficiencies in low-salinity water, researchers have examined relationships between ratios of metabolically important ions and growth and survival. Red drum survival was positively correlated to Na\(^{+}:K^{+}\) molar ratio and negatively correlated to K\(^{+}:Cl^{-}\) molar ratio in test water at 15 g L\(^{-1}\) salinity (Forsberg et al. 1996). However, the authors attributed these correlations to high within-treatment variation. Fielder et al. (2001) evaluated short-term growth and survival of Australian snapper in brackish ground water (19.6 g L\(^{-1}\) salinity) amended with potassium chloride (K\(^{+}:Cl^{-}\) molar ratio varied from 0.001–0.018). Fish died at a K\(^{+}:Cl^{-}\) ratio less than 0.007 and maximum growth occurred above a ratio of 0.010. Juvenile barramundi (Lates calcarifer [Bloch]) died during a bioassay in saline groundwater whose K\(^{+}:Cl^{-}\) molar ratio was 0.005 (Partridge & Creeper 2004). The principal cause of barramundi death, as determined by post-mortem examination, was skeletal myopathy caused by potassium-deficient groundwater.

Seven-d shrimp survival was not correlated with Na\(^{+}:K^{+}\), Ca\(^{2+}:Mg\(^{2+}\), or Na\(^{+}:Ca^{2+}\) molar ratios in environments where salinity (1–5 g L\(^{-1}\)) was derived from sea salt and mixed salts dissolved in soft water (Atwood et al. 2003). However, there was a positive correlation between survival and K\(^{+}:Cl^{-}\) molar ratio over the range of 0.008–0.018. Shrimp survival at 21 days was not correlated to any ion ratio. In another study at 30 g L\(^{-1}\) salinity, no L. vannamei survived at a Na\(^{+}:K^{+}\) molar ratio of 187.4, and the growth and energy budget of shrimp in water with a Na\(^{+}:K^{+}\) ratio of 153.3 was significantly lower than for shrimp at lower ratios (Zhu et al. 2004).

In the present experiment, the Na\(^{+}:K^{+}\) and Na\(^{+}:Ca^{2+}\) molar ratios were lower than those in seawater at the same salinity because concentrations of potassium and calcium were relatively higher and concentration of chloride was relatively lower. Molar ratios of K\(^{+}:Cl^{-}\) and Ca\(^{2+}:Mg\(^{2+}\) were higher than those in seawater at the same salinity because concentration of chloride was relatively lower, and concentrations of calcium and magnesium were relatively higher. The excess or deficient concentration compared with dilute seawater at the same salinity of calcium (555%), magnesium (146%), potassium (334%), sulphate (130%), chloride (65%) or sodium...
(93%) did not appear to impact negatively shrimp survival and growth. Clearly, *L. vannamei* grew well in an ionic environment that differed from that of dilute seawater. Perhaps achieving a minimum concentration of major ions is important to ensuring survival in ion-deficient waters. McGraw and Scarpa (2003) suggested that a minimum K\(^+\) concentration of 1 mg L\(^{-1}\) in a 1 g L\(^{-1}\) salinity water was required for *L. vannamei* survival. However, shrimp survival was monitored only for 48 h. It is unlikely shrimp would survive for an extended period in this water because the K\(^+\):Cl\(^-\) molar ratio was 0.0014. What remains is to determine the minimum concentrations of major ions for good survival and growth of *L. vannamei*.

Shrimp yields ranged from 3449 kg ha\(^{-1}\) in 2003 to 4966 kg ha\(^{-1}\) in 2004 in ponds stocked with 30–39 PL\(_{15}\) m\(^{-2}\) for a 125–134-d culture period. At harvest, mean individual weight ranged from 17.1–19.3 g shrimp\(^{-1}\), which were considered marketable size shrimp. Similar results have been reported for *L. vannamei* in studies in coastal ponds. During a 112-day experiment where PL\(_{25}\) *L. vannamei* was stocked at 35 PL m\(^{-2}\), mean yield was 3525 kg ha\(^{-1}\) and mean weight was 15.0 g shrimp\(^{-1}\) (Garza de Yta, Rouse & Davis 2004). The difference in mean weight between the two experiments likely resulted from a lower survival and higher growth rate in the present experiment. In another 112-day study stocked at 35 PL m\(^{-2}\), age of *L. vannamei* at stocking did not affect significantly mean gross yield (3375–4005 kg ha\(^{-1}\)) or mean individual weight at harvest (150–169 g shrimp\(^{-1}\)) (Zelaya, Rouse & Davis 2007). After a 4-month growing season in ponds on an inland farm stocked with 40 PL m\(^{-2}\), *L. vannamei* yield averaged 4500 kg ha\(^{-1}\) and mean individual weight ranged from 17.5–21.0 g shrimp\(^{-1}\) (Samocha et al. 2002). Mean *L. vannamei* yield after 203 days ranged from 2920–3325 kg ha\(^{-1}\) in ponds stocked with 15 PL m\(^{-2}\) on a farm in Sonora, Mexico (Casillas-Hernández, Magallon-Barajas, Portillo-Clarck & Paéz-Osuna 2006). However, because of the longer grow out period mean final weight (29.5–32.3 g shrimp\(^{-1}\)) exceeded that obtained in the present study.

While total shrimp yield for the growing season was similar for the PL\(_{15}\) and PL\(_{25}\) treatments, producing two crops of marketable shrimp during the early May to early October growing season by stocking PL\(_{25}\) shrimp appears doubtful because of the low mean weight at harvest. If ponds had been stocked in early May as opposed to late May, mean final weights would have been greater in both the PL\(_{15}\) and PL\(_{25}\)-first cycle treatments. However, it is unlikely the shrimp harvested from the PL\(_{25}\)-first cycle treatment would have been larger than those harvested from the PL\(_{25}\)-second cycle treatment.

In summary, this is the first published report that freshwater can be amended with key ions to permit production of *L. vannamei* in inland ponds. Salinity of source freshwater was increased to 0.7 g L\(^{-1}\). Thus, the presence of brackish ground water does not appear to be a necessary prerequisite for inland culture of *L. vannamei*. However, the ionic composition of source water will determine the quantities of minerals necessary to amend ion concentrations to desired levels. Additional research is needed to determine the minimum concentrations of major anions and cations for *L. vannamei*.

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**References**


