Efficacy of Pyriproxyfen for Control of Stored-Product Psocids (Psocoptera) on Concrete Surfaces

CHRISTOS G. ATHANASSIOU,1,2,3 FRANK H. ARTHUR,2 NICKOLAS G. KAVALLIERATOS,4 AND JAMES E. THRONE2

J. Econ. Entomol. 104(5): 1765–1769 (2011); DOI: http://dx.doi.org/10.1603/EC10424

ABSTRACT The insect growth regulator pyriproxyfen was evaluated as a surface treatment for control of three stored-product psocid pests Liposcelis bostrychophila Badonnel, Liposcelis decolor (Pearman), and Liposcelis paeta Pearman (Psocoptera: Liposcelididae). Nymphs were exposed for 35 d on a concrete surface treated with 2.3 mg of active ingredient/m² pyriproxyfen. Exposure to pyriproxyfen significantly reduced the numbers of both adults and nymphs in comparison with untreated controls. In adults, the greatest reduction (≥90%) was for L. decolor and L. bostrychophila, whereas for L. paeta it was 49%. Few adults of any species were found in the pyriproxyfen treatments. The greatest numbers of nymphs were recorded for L. bostrychophila for both pyriproxyfen treatments and controls. Few adults of any species were found in the pyriproxyfen treatments. The results indicate that pyriproxyfen is effective for control of L. bostrychophila, L. decolor, and L. paeta on concrete, and although complete control was not achieved, the results warrant further long-term study to determine whether pyriproxyfen can completely eliminate psocid populations over time.

KEY WORDS pyriproxyfen, insect growth regulator, Liposcelis, surface treatment, Psocoptera

Stored-product psocids (Psocoptera) belonging to the genus Liposcelis are important emerging pests of stored products. They feed on a wide variety of foods, but they are particularly associated with amylaceous commodities, such as grain and flour (Nayak 2006, Throne et al. 2006). One of the most important characteristics of these psocids is their high tolerance to many of the insecticides currently used for control of stored-product beetles (Nayak et al. 1998). Moreover, the bacterium-derived grain protectant spinosad was found to be ineffective against several Liposcelis spp. (Nayak et al. 2003, Athanassiou et al. 2009). Several organophosphate, carbamate, and pyrethroid insecticides have been evaluated for control of stored-product psocids on concrete surfaces, but they failed to provide long-term protection (Collins et al. 2000; Nayak et al. 2002a,b, 2003).

Insecticides that are registered to control stored-product beetles should be evaluated because of the increasing pest status of psocids. One such insecticide is the newly registered insect growth regulator (IGR) pyriproxyfen, which is labeled for use as a surface or spot treatment in food facilities in the United States. Several studies have documented its high efficacy against stored-product insect pests. Arthur et al. (2009) reported that pyriproxyfen was superior to hydroprene for control of several insect pests on wood, metal, and concrete surfaces, with a considerable level of persistence. Kostyukovsky et al. (2000) reported that pyriproxyfen was able to control pirimiphos-methyl-resistant Tribolium castaneum (Herbst) (Coleoptera: Tenebrionidae) populations and that it was superior to methoprene for control of the rice weevil, Sitophilus oryzae (L.) (Coleoptera: Curculionidae), and the lesser grain borer, Rhyzopertha dominica (F.) (Coleoptera: Bostrycidae). Hubert et al. (2007) found that a combination of permethrin + benzyl benzoate + pyriproxyfen was superior to chlorpyriphos, deltamethrin, beta-cyfluthrin, and a combination of deltamethrin + bioallethrin for control of several stored-product mites. However, there are no data available on efficacy of pyriproxyfen for control of stored-product psocids. The objective of our study was to determine whether pyriproxyfen applied at the label rate would be an effective surface treatment for control of psocids.
Materials and Methods

Insects. Young nymphs ($N_1-N_2$) of the psocids *Liposcelis bostrychophila* Badonnel, *Liposcelis decolor* (Pearman), and *Liposcelis paeta* Pearman (Psocoptera: Liposcelididae) were used in the tests. *L. bostrychophila* generally is parthenogenetic (males have only recently been discovered; Mockford and Krushelnicky 2008), whereas the other two species reproduce after mating. Psocids were reared on a mixture of 97% cracked wheat kernels, 2% crisp rice, and 1% brewer’s yeast at 30°C and 70% RH. To obtain individuals of standardized age for experiments, 30 female adults were left to oviposit in 35-mm petri dishes containing red-colored psocid diet for 3 d, and then the females were removed.

After an additional 7 d, the newly emerged nymphs were removed with a fine brush (Opit and Throne 2008). The colored diet was prepared by mixing 100 g of crisp rice with a solution of 5 ml of red food dye (Global ChemSources, Inc., Cedar Grove, NJ) in 300 ml of water. Afterward, the mixture was dried in a mechanical convection oven (HTM 85, Precision Scientific, Inc., Chicago, IL) for 2 d, and then the dried mixture that resulted was grinded in a Wiley Mill by using a #20 sieve (0.85-mm openings; Scientific Apparatus, Philadelphia, PA).

Insecticide. A formulation of pyriproxyfen (Nyguard, McLaughlin Gormley King [MGK] Company, Minneapolis, MN) that contains 10% active ingredient (AI) was used in the experiments at the label rate of 2.3 mg (AI)/m².

Bioassays. All tests were conducted in petri dishes (9 cm in diameter by 1.5 cm in height), which had a surface area of $\approx 62$ cm². The bottoms of the dishes were covered with driveway patching material (Rockite, Hartline Products Co., Inc., Cleveland, OH) to create the concrete surface (hereby referred to as arenas). The arenas were prepared 1–2 d before the tests. The internal sides of the arenas were coated with Fluon (polytetrafluoroethylene, Northern Products, Woonsocket, RI) to prevent escape of psocids. The solutions of pyriproxyfen were prepared with distilled water. Dishes were sprayed with a Badger 100 artist’s airbrush (Badger Corporation, Franklin Park, IL) to treat each individual concrete arena with 0.25 ml of formulated solution per 62 cm² area of the arena, which is equivalent to the label spray rate 3.81/94 m². Twenty-four arenas were prepared for each psocid species: 12 were sprayed with pyriproxyfen, and 12 were sprayed with water as untreated controls. Ten psocids and five kernels of cracked wheat were placed on each arena, either before or after spraying. The 24 arenas were divided into four categories (six replicate dishes per category): 1) arenas that contained psocids before spraying, with the kernels added after spraying; 2) arenas that contained psocids and kernels before spraying; 3) arenas that did not contain psocids before spraying (the psocids were added immediately after spraying) but contained five cracked kernels of wheat; and 4) arenas that did not contain psocids or kernels before spraying (the psocids and the kernels were added immediately after spraying).

This procedure was replicated three times, with new arenas each time (3 by 24 = 72 dishes for each species). After spraying, all arenas were placed in black plastic boxes containing saturated sodium chloride solution below a false floor to maintain the relative humidity at $\approx 75\%$. The boxes were placed in incubators maintained at 30°C and 70% RH. All arenas were checked after 35 d of exposure, and the numbers of adults and nymphs were recorded separately for each arena.

Data Analysis. For each species, the numbers of adults and nymphs in the treated concrete arenas were compared with those in the respective control dishes using a two-tailed $t$-test with $n$-2 df to indicate whether pyriproxyfen affected the number of psocids. Then, for each species and life stage, the data from the pyriproxyfen-treated dishes were analyzed using a one-way analysis of variance (ANOVA) to determine differences in number of psocids among the four treatment categories with JMP software (Sall et al. 2001). Means were separated by the honestly significant difference (HSD) test at $\alpha = 0.05$ (Sokal and Rohlf 1995).

Results

Adults. Numbers of adults were significantly reduced for all species in the pyriproxyfen-treated arenas compared with the numbers of adults in the controls (Table 1). The greatest reduction ($>90\%$) was for *L. decolor* and *L. bostrychophila*. For these species, less than one adult per arena was recorded. The number of *L. paeta* adults was reduced by only 49%. Significantly more *L. decolor* adults were recorded in arenas in which the kernels were treated first and the nymphs added afterward in comparison with the arenas in which the psocids were treated first and the

<table>
<thead>
<tr>
<th>Treatment</th>
<th><em>L. decolor</em> Adults</th>
<th><em>L. decolor</em> Nymphs</th>
<th><em>L. paeta</em> Adults</th>
<th><em>L. paeta</em> Nymphs</th>
<th><em>L. bostrychophila</em> Adults</th>
<th><em>L. bostrychophila</em> Nymphs</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>9.4 ± 1.0a</td>
<td>6.9 ± 1.5a</td>
<td>3.9 ± 0.7a</td>
<td>4.7 ± 1.7a</td>
<td>8.5 ± 0.3a</td>
<td>60.1 ± 5.3a</td>
</tr>
<tr>
<td>Pyriproxyfen</td>
<td>0.9 ± 0.2b</td>
<td>2.9 ± 1.2b</td>
<td>2.0 ± 0.4b</td>
<td>0.9 ± 0.6b</td>
<td>0.6 ± 0.3b</td>
<td>11.1 ± 4.2b</td>
</tr>
<tr>
<td><em>t</em></td>
<td>$&lt;0.01$</td>
<td>0.02</td>
<td>0.01</td>
<td>0.02</td>
<td>$&lt;0.01$</td>
<td>$&lt;0.01$</td>
</tr>
<tr>
<td><em>P</em></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Within each column, means followed by the same letter are not significantly different (df = 70; two-tailed $t$-test).
kernels added afterward or with arenas which contained psocids and kernels before spraying (Table 2). For *L. paeta* and *L. bostrychophila*, significantly higher numbers of adults were in arenas that did not contain psocids or kernels before spraying in comparison with the arenas in which the kernels were treated first and the psocids added afterward or vice versa (Tables 3 and 4).

**Nymphs.** The numbers of nymphs in the controls were significantly greater than the numbers of nymphs in the pyriproxyfen-treated dishes (Table 1). For both pyriproxyfen-treated and control arenas, the greatest number of nymphs was recorded for *L. bostrychophila*. Significantly more *L. decor* nymphs were recorded in arenas that contained only kernels before spraying, in comparison with other treatments (Table 2). In contrast, no significant differences were noted in numbers of *L. paeta* nymphs among the four treatments (Table 3), and nymphs were found only in arenas that contained psocids and kernels before spraying. Significantly more *L. bostrychophila* nymphs were in arenas that did not contain psocids or kernels before spraying in comparison with the arenas in which the kernels were treated first and the psocids added afterwards or vice versa (Table 4).

**Discussion**

This is the first study in which the IGR pyriproxyfen was evaluated on concrete surfaces for control of stored-product psocids, and this is also one of the few studies that examined this insecticide for control of stored-product insects in general. Our results indicate that pyriproxyfen can reduce numbers of psocids on concrete but that the level of reduction is mainly dependent on the target species. There have been several studies on another IGR, methoprene, for control of psocids. In a recent study, Athanassiou et al. (2010b) found that methoprene was not able to control the psocids *L. decor*, *L. bostrychophila*, *Liposcelis entomophila* (Enderlein), and *L. paeta* on maize (*Zea mays* L.), rice (*Oryza sativa* L.), and wheat (*Triticum aestivum* L.). Also, the combination of methoprene with spinosad was not effective for control of *L. bostrychophila* on wheat (Athanassiou et al. 2010a). These studies and earlier studies indicate lack of efficacy of methoprene for control of *Liposcelis* spp. (Nayak et al. 1998, 2002b). Bucci (1994) reported that *L. bostrychophila* was extremely tolerant to the IGR fenoxycarb, even at very high doses. These studies and our current findings indicate that psocids generally are quite tolerant to IGRs, including pyriproxyfen, although pyriproxifen is the most effective of IGRs evaluated thus far.

Access to food can moderate the efficacy of a given insecticide (Arthur 1998, 2009; Arthur et al. 2009). Arthur (2009) noted that survival of *T. castaneum* adults on concrete treated with chlorfenapyr increased when flour was present. Consequently, sanitation (presence or absence of food material) is an important variable that should be incorporated into evaluation of insecticides for stored-product pests. In previous studies, food was usually added after the application of the insecticide; thus, food was not treated. This may have allowed insects to moderate the toxic effects of the insecticides through feeding on untreated food. In our study, both cases were examined: insecticide-treated and insecticide-free food. There were no general trends in survival for either of food categories for all three species. We hypothesize that the presence of cracked kernels in all dishes might have reduced pyriproxyfen efficacy, although treatments without kernels were not evaluated in this study. These results suggest that removal of insecticide-treated food residues in a milling or warehouse facility may be just as important for pest control as removal of untreated food residues.

Differences among psocid species in their susceptibility to insecticides applied on concrete is an additional parameter that should be taken into consider-

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Adults</th>
<th>Nymphs</th>
</tr>
</thead>
<tbody>
<tr>
<td>Without psocids, without kernels</td>
<td>4.1 ± 1.2a</td>
<td>0.0 ± 0.0a</td>
</tr>
<tr>
<td>Without psocids, with kernels</td>
<td>1.1 ± 0.8bc</td>
<td>0.0 ± 0.0a</td>
</tr>
<tr>
<td>With psocids, without kernels</td>
<td>0.0 ± 0.0c</td>
<td>0.0 ± 0.0a</td>
</tr>
<tr>
<td>With psocids, with kernels</td>
<td>1.9 ± 0.6ab</td>
<td>3.7 ± 2.5a</td>
</tr>
<tr>
<td><em>F</em></td>
<td>6.2</td>
<td>2.3</td>
</tr>
<tr>
<td><em>P</em></td>
<td>&lt;0.01</td>
<td>0.10</td>
</tr>
</tbody>
</table>

Within each column, means followed by the same letter are not significantly different (df = 3, 32; HSD test at 0.05; “without” or “with” indicate absence or presence during spraying, respectively).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Adults</th>
<th>Nymphs</th>
</tr>
</thead>
<tbody>
<tr>
<td>Without psocids, without kernels</td>
<td>2.0 ± 0.5a</td>
<td>30.0 ± 10.5a</td>
</tr>
<tr>
<td>Without psocids, with kernels</td>
<td>0.0 ± 0.0b</td>
<td>3.4 ± 3.0bc</td>
</tr>
<tr>
<td>With psocids, without kernels</td>
<td>0.0 ± 0.0b</td>
<td>0.1 ± 0.1c</td>
</tr>
<tr>
<td>With psocids, with kernels</td>
<td>0.4 ± 0.4ab</td>
<td>10.7 ± 7.7db</td>
</tr>
<tr>
<td><em>F</em></td>
<td>4.3</td>
<td>3.9</td>
</tr>
<tr>
<td><em>P</em></td>
<td>0.01</td>
<td>0.04</td>
</tr>
</tbody>
</table>

Within each column, means followed by the same letter are not significantly different (df = 3, 32; HSD test at 0.05; “without” or “with” indicate absence or presence during spraying, respectively).
ation when evaluating insecticides. Among the species tested here, *L. paeta* is considered to be tolerant to several insecticides, such as chlorpyrifos-methyl, pirimiphos-methyl, and fenitrothion, whereas *L. bostrychophila* is considered susceptible (Collins et al. 2000, Nayak et al. 2002a). In our study, there were considerably greater numbers of *L. bostrychophila* nymphs, in both treated and untreated dishes, in comparison with the other two species. For example, the number of *L. bostrychophila* nymphs was 13 and 7 times higher than the numbers of *L. paeta* and *L. decolor* nymphs, respectively. In contrast with the other two species, *L. bostrychophila* typically reproduces parthenogenetically, with high potential for production of progeny (Wang et al. 2000). Athanassiou et al. (2010c) found that *L. bostrychophila* adult females generally produced considerably more F1 progeny than *L. paeta* or *L. decolor* in a wide variety of grain commodities. Even in insecticide-treated wheat and rice, Athanassiou et al. (2009) reported that *L. bostrychophila* produced greater numbers of offspring than *L. paeta*. Greater capacity for progeny production may counteract the insecticidal effect, especially in the case of IGRs, where this effect is primarily focused on the immature life stages. Nevertheless, pyriproxyfen reduced *L. bostrychophila* numbers in comparison with the control, but the level of reduction varied according to the treatment. In a previous study on wheat, rice, and maize, methoprene did not effectively control *L. bostrychophila*, *L. decolor*, or *L. paeta*, although that progeny production was reduced for all species.

Few adults of any species were found at the end of the 35-d exposure interval. It is well documented that 35 d is a sufficient interval for young psocid nymphs to reach the adult stage at the conditions examined here (Bees and Walker 1990, Leong and Ho 1995, Wang et al. 2000, Opit and Throne 2008). Therefore, we can conclude that there is an effect of pyriproxyfen on the species tested. This was evident even in the case of *L. bostrychophila* where the greatest number of nymphs was recorded on the treated substrates, yet the lowest number of adults was found. The exposure interval was selected to examine whether the exposed nymphs would reach the adult stage and the capacity of these adults for progeny production. However, we do not know whether the nymphs found on the pyriproxyfen-treated surfaces had been arrested at that stage and were unable to become adults. For *Plodia interpunctella* (Hubner) (Lepidoptera: Pyralidae), Mohandass et al. (2006) found that hydroprene prolonged immature development or caused molting failure. Arthur et al. (2009) found that pyriproxyfen reduced adult emergence of the cigarette beetle, *Lasioderma serricorne* (F.) (Coleoptera: Anobiidae); the confused flour beetle, *Tribolium confusum* Jacqueulin du Val (Coleoptera: Tenebrionidae); the sawtoothed grain beetle, *Oryzaephilus surinamensis* (L.) (Coleoptera: Silvanidae); *T. castaneum*; and *P. interpunctella* after 56 d of exposure, whereas, at the same conditions, hydroprene lost its effectiveness after 28 d. Testing longer exposure intervals would indicate whether the activity of pyriproxyfen can gradually eliminate the psocid population.

In conclusion, pyriproxyfen showed activity against *L. decolor*, *L. paeta*, and *L. bostrychophila* on concrete, a characteristic that should be further evaluated to determine whether pyriproxyfen can eliminate psocid populations over time. In addition, the combined application of pyriproxyfen with a reduced-risk contact neurotoxic insecticide, such as chlorfenapyr (Guedes et al. 2008, Arthur 2009), may further increase its efficacy.

Acknowledgments

We thank Ann Redmon and Ngunza Kisangani for technical assistance, MGK for the pyriproxyfen sample, and Manoj Nayak for comments on an earlier version of this manuscript.

References Cited


Received 20 November 2010; accepted 30 April 2011.